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(54) Title: INTRACELLULAR ISOFORM OF THE INTERLEUKIN-I RECEPTOR ANTAGONIST

(57) Abstract

It is described a new interleukin-1 antagonist active both against IL-1a and IL-1B, a new DNA sequence encoding the IL-1 antagonist and the method for obtaining a IL-1 antagonist by the recombinant DNA technique; it is also described the prophylactic, therapeutic and diagnostic use of such new IL-1 antagonist in pathologies deriving from the IL-1 production.

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Intracellular isoform of the interleukin-1 receptor antagonist

FIELD OF THE INVENTION

The present invention is in the field of biotechnology. It is described a new interleukin-1 (IL-1) antagonist active both against IL-1a and IL-1B, a new DNA sequence encoding the IL-1 antagonist and the method for obtaining a IL-1 antagonist by the recombinant DNA technique. It is also described the prophylactic, therapeutic and diagnostic use of such new IL-1 antagonist in pathologies deriving from the IL-1 production.

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BACKGROUND OF THE INVENTION

There are two distinct genes encoding the interleukin-1 (IL-1) named IL-1a and IL-1B, which encode protein IL-1a and IL-1B respectively.

Interleukin IL-1a and IL-1B are pleiotropic cytokines, which, although their sequences show scarce analogy, exert a variety of similar effects on different tissues and act on many human pathologies, in particular on the immunitary response of the organism and on inflammatory processes.

Both the proteins have a molecular weight of about 17.5 KDa and have been previously synthesised as precursor molecule of larger size having a molecular weight of about 31 KDa.

IL-1s are potent inflammatory and pyrogenic cytokines that normally have beneficial effects but can also have extremely unhealthy effects for the organism.

They can, for example, participate in the pathogenesis of symptoms of the autoimmune pathologies like lupus eritematosus and, in particular, they are involved as mediators to provoke damages to tissues as for example in rheumatoid arthritis.

Many of the biological effects of IL-1 are similar to those that can be observed during a septic event. Recent studies demonstrated that the endovenous administration of IL-1 in doses from 1 to 10 ng/kg gives rise to fever, sleepiness, anorexia, generalised myalgia, arthralgia and cephalea.

Since IL-1 have pleiotropic biological activities, many of which influence negatively the organism, the powerful effects of IL-1 should be under strict physiological control.

IL-1 synthesis is inhibited by anti-inflammatory cytokines, prostaglandins and glucocorticoids and the existence of multiple levels of inhibition of IL-1 points to the necessity of a strict control of this mediator.

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IL-1 is the only cytokine for which an antagonist polypeptide for the receptor has been described up to now: the third known component until today of the IL-1 family is the antagonist for the IL-1 receptor (IL-1ra).

All three components (IL-1a, IL-1B, IL-1ra) recognise and bind to the same receptor on cell surface (IL-1R); IL-1a and IL-1B binding to IL-1R transmit a signal, whilst IL-ra does not.

There are two types of IL-1 receptors named IL-1RI and IL-1RII. IL-1ra is a polypeptide which binds IL-1RI, and with less affinity IL-1RII, without any agonistic activity.

IL-1ra production is induced in different cellular types, including mononuclear phagocytes, polymorphonuclear cells (PMN) and fibroblasts, by IgG, cytokines and bacterial products.

Until now two molecular forms of IL-1ra have been identified and cloned:

1) secreted IL-1ra (sIL-1ra) contains a classical leader sequence of 25 amino acids giving a mature protein of 152 amino acids; 2) intracellular IL-1ra (icIL-1ra) lacks a leader sequence thus predicting that this protein remains intracellular.

sIL-1ra and icIL-1ra are generated from the same gene. icIL-1ra transcripts originate from an alternative starting site and from the splicing of a first alternative exon into an internal splice acceptor site located in the first exon of sIL-1ra. The predicted proteins are thus identical except in their NH₂ ends, where the first 21 amino acids of sIL-1ra are substituted by four amino acids in icIL-1ra.

Expression of transcripts encoding sIL-1ra and icIL-1ra is differently regulated. The biological significance of icIL-1ra is still unclear.

Considering that IL-1 is involved in pathogenesis of many diseases it is evident the need of having available medicaments useful to limit the unhealthy effects of IL-1.

30 SUMMARY OF THE INVENTION

An object of the present invention is to provide an IL-1 antagonist active against both IL-1a and IL-1B and against a combination of them.

A further object of the present invention is to provide a DNA sequence encoding an IL-1 antagonist and a method for obtaining such new antagonist by the recombinant DNA technique.

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Another further object of the present invention is to provide the antagonist in substantially purified form in order to be suitable for use in pharmaceutical compositions active in pathologies that require IL-1 inhibition.

Further objects and advantages of the invention will be evident in the following description.

BRIEF DESCRIPTION OF THE FIGURES AND SEQUENCE LISTING

Figure 1 describes the DNA sequence and the sequence of the protein, for the portion not in common, of icIL-1raII (SEQ ID NO:8 and SEQ ID NO:9) compared to those of classic sIL-1ra (icIL-1raI; SEQ ID NO:6) and of sIL-1ra (SEQ ID NO:4 and SEQ ID NO:5), and it further describes the DNA sequence and the encoded protein for the portion of IL-1ra in common (SEQ ID NO:13 and SEQ ID NO:14).

Figure 2 describes the RT-PCR analysis of icIL-1raII expression in different cell types.

Figure 3 describes the Western blot analysis of recombinant ic IL-1raII.

Figure 4 describes the effects of icIL-1raII on IL-1 induced expression of E-selectin in endothelial cells.

SEQ ID NO:1 reports the sequence of an oligonucleotide named IRA5 for use in RT-PCR.

SEQ ID NO:2 reports the sequence of an oligonucleotide, corresponding to nucleotides 69-70 of B-actin cDNA, for use in RT-PCR.

SEQ ID NO:3 reports the sequence of a backward oligonucleotide, complementary to nucleotides 430-449, for use in RT-PCR.

SEQ ID NO:4 reports the DNA sequence encoding sIL-1ra for the portion not in common.

SEQ ID NO:5 reports the amino acid sequence of sIL-1ra for the portion not in common.

SEQ ID NO:6 reports the DNA sequence encoding three amino acids of icIL-1ral for the portion not in common.

SEQ ID NO:7 reports the three amino acids of icIL1-ral for the portion not in common.

SEQ ID NO:8 reports the DNA sequence encoding icIL-1raII for the portion not in common.

SEQ ID NO:9 reports the amino acid sequence of icIL-1raII for the portion not in common.

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SEQ ID NO:10 reports the DNA sequence encoding IL-1ra for the portion in common. With regard to questions related to the "Patentin EPO" program, for the preparation of the sequences a G nucleotide was added in the first position of the sequence in order to permit the encode of the first amino acid Glu and further in order to avoid the formation of a stop codon in the inner side of the sequence.

SEQ ID NO:11 reports the amino acid sequence of IL-1ra for the portion in common.

SEQ ID NO:12 reports the sequence of 21 amino acids representing a icIL-1raII fragment not in common with the other IL-1ras.

SEQ ID NO:13 reports the DNA sequence encoding the complete icIL-1raII.

SEQ ID NO:14 reports the amino acid sequence of complete icIL-1raII.

15 DESCRIPTION OF THE INVENTION

This new IL-1 antagonist was generated by inserting in the frame of the DNA encoding icIL-1ra a new 63 base pairs (bp) sequence between the first icIL-1ra specific exon and the internal acceptor site of the first exon of sIL-1ra.

By RT-PCR experiments the present inventors found that this new transcript is expressed in activated monocytes and fibroblasts and in polymorphonuclear cells (PMN).

Expression in COS cells revealed that this new antagonist is mostly intracellular and has a molecular weight (MW) of approximately 25 KDa in SDS-PAGE.

The new recombinant antagonist shows IL-1 inhibitory activity.

In the present application, for reason of clearness and easiness, the presently known icIL-1ra are indicated as icIL-1ra type I (icIL-1raI), whereas the new antagonist here described and object of the present invention is defined as icIL-1ra type II (icIL-1raII).

Examples of pathologies in which the new antagonist according to the invention can be advantageously used for prophylactic, therapeutic or diagnostic use are rheumatoid arthritis, septic shock, acute myelomonocytic leukaemia, immunological reaction of transplantation against host, acquired immunodeficiency syndrome (AIDS), ulcerative colitis and all autoimmune diseases in general.

An embodiment of the invention is the administration of a pharmacological active amount of icIL-1raII to people having a high risk to

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develop pathologies requiring IL-1 inhibition or to people already showing pathologies like sepsis.

An example of the category above cited are patients waiting for a surgical operation.

Any route of administration compatible with the active principle can be used, but particularly preferred is the parenteral administration because it permits to have, in short times, systemic effects.

For this reason, it is preferable the administration of a endovenous bolus just before, during or after the surgical operation. The dose of icIL-1rall to be administered depends on the basis of the medical prescriptions according to age, weight and the individual response of the patient.

The dosage can be between 0.05 and 30 mg/Kg body weight and the preferable dose is between 0.1 and 10 mg/Kg body weight.

The pharmaceutical composition for parenteral use can be prepared in injectable form comprising the active principle and a suitable vehicle. Vehicles for the parenteral administration are well known in the art and comprise, for example, water, saline solution, Ringer solution and dextrose.

The vehicle can contain smaller amounts of excipients in order to maintain the solution stability and isotonicity.

The preparation of the cited solutions can be carried out according to the ordinary modalities and preferably the icIL-1raII content will be comprised between 1 mg/ml and 10 mg/ml.

Further examples of pathologies wherein the new antagonist according to the invention can be advantageously used for prophylactic, therapeutic diagnostic purpose are rheumatoid arthritis, septic shock, acute myelomonocytic leukaemia, immunological reaction of transplantation against host, acquired immunodeficiency syndrome (AIDS), ulcerative colitis and all autoimmune diseases in general.

The present invention has been described with reference to the specific embodiments, but the content of the description comprises all modifications and substitutions which can be brought by a person skilled in the art without extending beyond the meaning and purpose of the claims.

In the following part some methods for obtaining the invention will be described, although equivalent materials and methods can be used. The following examples are therefore purely illustrative and non-limiting of the invention.

EXAMPLE 1

Cloning and characterisation of icIL-1raII

MATERIALS AND METHODS

Reagents

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The following commercially available reagents were used for culture and separation of cells: pyrogenfree saline and distilled water for clinical use; RPMI 1640 medium; DMEM medium; M199 medium; L-glutamine; Percoll; Ficoll-Hipaque; aseptically collected fetal calf serum; endothelial cells growth supplement (ECGS), prepared from bovine brain; Heparin.

All reagents contained less than 0.125 EU/ml of endotoxin as checked by thr Limulus amebocyte lysate assay.

Cells

Human circulating PMN and monocytes were separated from the peripheral blood of healthy donors by centrifugation on a discontinuous (46% for monocytes and 62% for PMN) gradient of isoosmotic (285 mOsm) Percoll, as described in Colotta F., Peri G., Villa SA., Mantovani A., Rapid killing of actinomycin D treated tumour cells by human mononuclear cells. J. Immunol. 132:936, 1984. Cells were recovered at the interface, washed twice in saline and resuspended in the medium.

PMN and monocytes recovery was higher than 90% and purity higher than 98%, as assessed by morphological examination of stained cytocentrifuged cells. The cell culture medium routinely used for PMN and monocytes was RPMI 1640 with 2 mM L-glutamine and 10% FCS.

Human endothelial cells (EC) were obtained from umbilical veins and cultured, as described in detail in the literature (Allavena P., Paganin C., Martin-Padura I., Peri G., Gaboli M., Dejana E., Marchisio P.C., Mantovani A., Molecules and structures involved in the adhesion of natural killer cells to vascular endothelium, J. Exp. Med., 173:439, 1991).

Confluent cells at 2nd-5th passage maintained in M199 medium with 10% FCS supplemented with ECGS (50 μ g/ml) and Heparin (100 μ g/ml) were routinely used.

COS cells were cultivated in DMEM medium with 10% FCS and 8387 fibroblast cells in RPMI 1640 medium with 10% FCS.

After the appropriate treatment, cells were examined for IL-1ra mRNA or IL-1ra protein as described below.

RT-PCR

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Total RNA was extracted by the guanidinium isothiocianate method with minor modifications.

RT-PCR was performed as described in Colotta F., Polentarutti N., Sironi M., Mantovani A., J. Biol. Chem., 267:18278, 1992.

Briefly, 1 µg total RNA was reverse transcribed in reverse transcriptase buffer (5 mM MgCl₂, 50 mM KCl, 10 mM Tris-HCl; pH 8.3) with 2.5 mM random hexamers, 1 mM each deoxynucleotide triphosphate, 1 unit/ml RNase inhibitor, and 2.5 units/ml moloney murine leukaemia virus transcriptase (Perkin Elmer Cetus, Norwalk, CT).

Samples were incubated for 10 min at 25°C and then at 42°C for 45 min. Then, cDNA reaction was added with a specific pair of primers designed to amplify cDNAs encoding icIL-1raI or icIL-1raII and, as an internal control, human B-actin.

Amplification was carried out in 2 mM MgCl₂, 50 mM KCl, 0.2 M each deoxynucleotide triphosphate, 2.5 units/100 ml Taq polymerase (Perkin Elmer Cetus) and 4 mg/ml of each specific primer (see below). Amplification (30 cycles) was carried out in an automated thermal cycler (Perkin Elmer Cetus) at 95°C, at 55°C and at 72°C for 1.5 min each.

Amplified products were run through a 1% ethidium bromide-stained agarose gel along with molecular weight standards (Boehringer Mannheim, Mannheim, Germany).

Oligonucleotides were synthesised by the phosphoramidite method. The sequences of oligonucleotides used to selectively amplify icIL-1ra were identical to those described in Haskill S. et al., Natl. Acad., USA, 88:3681, 1991.

In particular, the authors used oligonucleotides GM397 (indicated here as IRA 1) and GM368 (IRA 4).

For icIL-1raII amplification the authors used IRA 4 and IRA 5 (SEQ ID NO:1), which specifically recognises the extra exon described here included in the icIL-1raII sequence.

For B-actin amplification the forward oligonucleotide is reported in SEQ ID NO:2, corresponding to nucleotides 60-79 of B-actin cDNA.

The backward oligonucleotide is reported in SEQ ID NO:3, complementary to nucleotides 430-449. Amplification products were subcloned (TA Cloning System, Invitrogen, San Diego, CA) and sequenced by the dideoxi chain termination method.

Expression of icIL-1ra products in COS cells

The cDNAs containing 32 bp of the 5'-untraslated region, the complete open reading frame and 6 bp (including the stop codon) of the 3'-untraslated region of both the icIL-1ral and icIL-1rall were obtained by RT-PCR with oligonucleotides IRA 4 and IRA 5 as detailed above and then ligated back into the pSF5 expression vector. Fidelity of reverse transcription and amplification was verified by sequencing.

The plasmids containing the cDNA in the correct orientation were purified on CsCl gradient and then transfected into COS cells by the calcium precipitate method as described in Sambrook J. et al., Cold Spring Harbor Laboratory Press, 1989.

After two days, culture supernatants and sonicated cell lysates examined by ELISA or immunoblotting as detailed below. An empty plasmid (not transfected) was used as a control.

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Identification of immunoreactive IL-1ra

A commercial ELISA test (Amersham, Buckinamshire, UK) that identifies both sIL-1ra and icIL-1ra was used. For the Western blot analysis polyclonal antisera of two rabbits and of one goat were used.

COS cells lysates samples and supernatants were run on 12.5% SDS-PAGE electrophoresis and then blotted onto a nitro-cellulose filter (Stratagene, La Jolla, CA, USA).

Incubation with primary and secondary antibodies was carried out according to standard protocols. The primary antibody was an anti-IL-1ra rabbit polyclonal antibody.

The secondary antibody was a goat anti-rabbit immunoglobulin fraction linked to horseradish peroxidase (Amersham). Immunoreactive protein fraction bands were revealed by a chemiluminescence-based procedure (ECL Detection, Amersham) according to manufacturer's instructions.

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Il-1-induced expression of E-selectin on EC

Confluent EC cultivated in 96 well plates (Falcon) were incubated for 30 minutes with an amount of transfected COS cells lysate (see above) corresponding to 25 to 100 ng of recombinant IL-1ra (either icIL-1ral or icIL-1raII) as assessed by a specific ELISA assay (Amersham).

As a control, an equal amount of COS lysate obtained from mock transfected cells was used in parallel. Next, EC were exposed for 6 hours to

0.1-1 ng/ml human recombinant IL-1B. The detection of E-selectin expression was made with an ELISA assay on adherent EC with the anti-E-selectin monoclonal antibody BB1G-E2 as primary antibody and a rabbit anti-mouse Ig antiserum conjugated with horseradish peroxidase as a secondary antibody. O.D. of the samples was determined by detecting the plates with a spectrophotometer (Flow) at 405 wavelength.

RESULTS

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Identification of icIL-1raII

Specific oligonucleotide primers were designed (indicated as IRA 1 and IRA 4 in Fig.1) in order to obtain the whole coding sequence of icIL-1ra (Fig.1) by RT-PCR. Amplified products from human PMN were subcloned and sequenced.

In addition to the previously known sequence of icIL-1ra, the inventor isolated a number of clones whose sequences were identical to the published icIL-1ra coding sequence, with the notable exception of an extra sequence of 63 bp between nucleotides 132 and 133 of the icIL-1ra sequence. Given the described exon-intron boundaries of icIL-1ra, the extra sequence is inserted between the first leader-less exon of icIL-1ra and the internal acceptor site of the first exon of sIL-1ra (Fig.1).

The predicted amino acid sequence is shown in Fig.1. The new protein (thereafter referred to as icIL-1ra type II) has the first three amino acids at the NH₂ terminus in common with the classical icIL-1ra (icIL-1ra type I), followed by a new sequence of 21 amino acids. The rest of the two proteins is identical.

Curiously, the junction with the internal acceptor site of the first exon of sIL-1ra always generated, both for sIL-1ra and icIL-1ral and for icIL-1rall, the same amino acid residue, i.e. glutamic acid (Fig.1).

The most striking characteristic of the inserted amino acid extra sequence is the presence of seven glycin residues, six of which are consecutive. Glycin residues are flanked on both sides by glutamic acid residues. icIL-1raII consists of 180 amino acids.

The overall hydrophilic pattern of icIL-1raII is similar to that of icIL-1raI, still lacking an hydrophobic leader peptide at the NH₂ terminus.

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Expression of icIL-rall

To identify icIL-1raII transcripts, RT-PCR analysis was performed with a pair of specifically designed oligonucleotides (IRA 5 and IRA 4, Fig.1), with an expected amplified product of 33 bp.

As shown in Fig.2, transcripts encoding icIL-1raII were detectable in PMA-, IL-1- and TNF-activated fibroblasts. A faint but detectable band was evident in LPS-treated monocytes.

Also PMN, either untreated or activated (Fig.2) showed a very faint band of the expected size.

The specificity of amplified products indicated in Fig.2 was confirmed by subcloning and sequencing.

Expression of recombinant icIL-1rall

COS cells were transfected with the DNA sequence encoding icIL-1raII and, by way of comparison, with that encoding icIL-1raI. Next, cell lysates and supernatants were examined by Western blot.

The polyclonal antisera used in these experiments recognised equally well icIL-1raII and icIL-1raI (Fig.3). Most, if not all, of icIL-1raII and icIL-1raI were found in cell lysates.

Recombinant icIL-1ral migrated as a predominant band of 22 KDa, whereas icIL-1rall showed a mass of approximately 25 KDa.

Inhibition of IL-1B activity by recombinant icIL-1rall

Recombinant ic IL-1 rall was examined for IL-1 inhibiting activity. To this aim the authors chose the IL-1-induced expression of E-selectin on endothelial cells, because this assay is sensitive (detectable induction at 100 pg/ml IL-1, or less) and rapid (6 hours incubation with IL-1).

Lysates of mock transfected COS cells did not significatively reduce the IL-1 activity.

icIL-1raII had no agonistic activity.

As shown in Fig.4, recombinant icIL-IraII inhibited in a dose-dependent fashion IL-1 activity.

These data provide evidence that ic IL-1 rall is indeed an inhibitor of IL-1

DISCUSSION

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The inventors describe a new molecular form of icIL-1ra. The new molecule is generated by insertion of 63 bp between the first leader-less exon of icIL-1ra and the internal acceptor site of the first exon of sIL-1ra.

Since the resulting protein is partially identical to classical icIL-1ra, with the exception of an extra sequence of 21 amino acids located in the NH₂ terminus of the molucule, the inventors suggest to term this new form as IL-1ra type II, referring to the classical icIL-1ra sequence as icIL-1ra type I.

RT-PCR experiments demonstrated that icIL-1raII transcripts are inducible in monocytes and fibroblasts. Recombinant icIL-1raII expressed in COS cells had an apparent MW of approximately 25 KDa and an inhibitor activity of IL-1 comparable to that exerted by icIL-1raI expressed under the same experimental conditions.

Transcripts coding for icIL-1ra and sIL-1ra are generated from the same gene by means of usage of differential splicing. icIL-1ra is generated by an alternative start of transcription of an exon inserted into an internal acceptor site of the first exon containing the leader sequence of sIL-1ra.

The results obtained by the inventors suggest a new organisation of IL-1ra gene, in which an extra exon is located between the first exon of, respectively, classical icIL-1ra and sIL-1ra. Use of this new exon generates a polypeptide molecule which, still lacking a signal peptide, differs from icIL-1ral at its N terminus by the insertion of 21 amino acids, still remaining inhibitory capacity against IL-1.

Use of alternative splicing to generate different IL-1ra molecules appears to be highly regulated. icIL-1raII transcripts were induced by IL-1, TNF and phorbol esters in fibroblasts and by LPS in monocytes. In fibroblasts, phorbol esters were found to selectively induce icIL-1ra transcripts, whereas IL-1 and TNF induced both sIL-1ra and icIL-1ra mRNAs. In monocytes, IL-13, which augmented both transcripts of sIL-1ra and icIL-1raI, failed to induce icIL-1raII.

Finally, PMN, in which sIL-1ra and icIL-1ra are costitutively expressed and inducible, expressed very few transcripts, as pointed out by RT-PCR. Overall, these data indicate that the mechanisms inducing the differential splicing generating the three forms of IL-1ra are differentially regulated in response to external signals.

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The amino acid sequence of the extra sequence described here is surprising in that it contains seven residues of glycin, six of which are consecutive.

Glycin-rich sequences are present in molecules with different biological activities, including the atrial natriuretic clearance receptor, the HOX11 home box gene, the intermediate filaments keratins and nuclear proteins involved in centromere binding or RNA splicing.

Apart from glycin residues, however, no obvious homology was evident between these proteins and icIL-1raII in the amino acid sequence flanking glycin-rich regions.

IL-1 system shows an extraordinary level of complexity, consisting of two agonists, two receptors, one of which is an inhibitor of IL-1, and a receptor antagonist, for which at least three different molecular forms could exist taking into account the results obtained.

Although the biological significance of the intracellular forms of IL-1ra remains to be clearly established, the data here reported indicate that by alternative splicing two different forms of icIL-1ra can be generated in response to selected external stimuli, with different N termini.

The existence of multiple and commex levels of control of IL-1 points to the absolute requirement for a tight physiological control of the inflammatory potential of this cytokine.

DESCRIPTION OF FIGURES

Figure 1

DNA sequence and predicted protein sequence of icIL-1raII compared to classical icIL-1ra(icIL-1raI) and sIL-1ra.

The upper part of Figure 1 shows DNA and protein sequences specifically represented in sIL-1ra, icIL-1ral and icIL-1raII. The lower part of Figure 1 shows the sequence in common among the three forms of IL-1ra.

The entire sequences for each molecule are thus generated by the junction of each specific portion with the common sequence. For clarity, the DNA sequence of icIL-1ra starts from nucleotide 91 of the published 5' untraslated sequence, and only 6 bp of the 3' untraslated sequence are reported.

The common IL-1ra sequence starts with the internal acceptor site located in the first exon of sIL-1ra, corresponding to nucleotide 133 of the complete icIL-1ral sequence and to nucleotide 88 of the complete sIL-1ra sequence.

Arrows indicate forward (IRA 1 and IRA 5) and backward (IRA 4) oligonucleotides used for RT-PCR analysis, as described in the text. The oligonucleotide IRA 5 recognises only icIL-1raII DNA.

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RT-PCR analysis of icIL-1raII expression in different cell types

RNAs from 8387 fibroblasts (panel A), monocytes (B) and PMN (C) were reverse-transcribed. Each DNA synthesis reaction was then divided in two samples, one of which amplified with oligonucleotides IRA 5 (forward) and IRA 4 (backward) for detection of icIL-1raII transcripts, and the other amplified with B-actin specific oligonucleotides (see Material and Methods Section).

Amplified products were then examined through an ethidium bromidestained agarose gel. Amplified products corresponding to B-actin are reported on the left side of the standard and the amplified products corresponding to icIL-1raII (on the right) are indicated by a arrow. The specificity of these bands was confirmed by subcloning and sequencing.

Figure 3

20 Western blot analysis of recombinant icIL-1raII

Cell lysates from COS cells transfected with DNAs encoding icIL-1raI (2) or icIL-1raII (3) or with an empty vector which does not contain such DNA (1) were examined by immunoblotting with an anti-IL-1ra rabbit polyclonal antibody. Molecular weight standards are indicated.

Figure 4

Effects of icIL-1raII on IL-1-induced expression of E-selectin on endothelial cells

Endothelial cells were treated with 0.1 or 1 ng/ml of human IL-1B, with or without 25-100 ng/ml of icIL-1raII or equivalent amounts of COS cell lysates obtained from cells which were mock transfected by means of an empty vector, as explained in details in the Material and Method section.

After 6 hours of incubation, the endothelial cells were examined for E-selectin expression by an ELISA test performed on adherent cells.

The data reported are percentages of IL-1-induced E-selectin expression for the control.

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SEQUENCE LISTING

- (1) GENERAL INFORMATION:
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 - (ii) TITLE OF INVENTION: INTERLEUKIN-1 ANTAGONIST
 - (iii) NUMBER OF SEQUENCES: 14
 - (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)
- (2) INFORMATION FOR SEQ ID NO: 1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (ix) FEATURE:
 - (A) NAME/KEY: misc_feature
 - (B) LOCATION:1..25
 - (D)OTHER INFORMATION:/note= "RT-PCR oligonucleotide named IRA5"
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

CTGACTTGTA TGAAGAAGGA GGTGG .

(2) INFORMATION FOR SEQ ID NO: 2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (ix) FEATURE:
 - (A) NAME/KEY: misc_feature
 - (B) LOCATION:1..20
 - (D) OTHER INFORMATION:/note= "RT-PCR oligonucleotide" corresponding to 60-79 of B-actin"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:
- GCGCTCGTCG TCGACAACGG

- (2) INFORMATION FOR SEQ ID NO: 3:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (ix) FEATURE:
 - (A) NAME/KEY: misc_feature
 - (B) LOCATION:1..21
- (D) OTHER INFORMATION:/note= "RT-PCR backward oligonucleotide complementary to 430-449"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

GATAGACAAC GTACATGGCT G

21

- (2) INFORMATION FOR SEQ ID NO: 4:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 87 base pairs

	(B)	TYPE: nucle	ic acid			
	(C)	STRANDEDNES	S: single			
	(D)	TOPOLOGY: 1	inear			
	(ii) MOLEC	CULE TYPE: C	DNA			
	(iii) HYPO	OTHETICAL: N	0			
	(iv) ANTI-	-SENSE: NO				
	(ix) FEAT	JRE:				
	(A)	NAME/KEY: C	DS			
	(B)	LOCATION:24	86			
	(ix) FEAT	URE:				
	(A)	NAME/KEY: m	isc_feature			
	(B)	LOCATION:1.	.87		`	
	(D)	OTHER INFOR	MATION:/note	= "Sequence	of sIL-lra not	in
		com	mon"			
			TION: SEQ ID			
GAATT	CCGGG CTGC.		TG GAA ATC TO			50
			et Glu Ile C	ys Arg Gly	Leu Arg Ser	
			1	5		
			TTC CTG TTC			87
His I	Leu Ile Thr	Leu Leu Leu	ı Phe Leu Phe	His Ser		
10		15		20		
(2)		FOR SEQ ID				
		NCE CHARACTE				
		LENGTH: 21				
		TYPE: amino				
	(D)	TOPOLOGY:	linear			
		CULE TYPE:				
			PTION: SEQ ID			
Met	Glu Ile Cys	Arg Gly Le			hr Leu Leu Leu	
1		5	10		15	
Phe	Leu Phe His	; Ser				
	20					
(2)	INFORMATION	N FOR SEQ ID	NO: 6:			

(i) SEQUENCE CHARACTERISTICS:

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-17-

- (A) LENGTH: 42 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 33..41
- (ix) FEATURE:
 - (A) NAME/KEY: misc_feature
 - (B) LOCATION:1..42
 - (D) OTHER INFORMATION:/note= "Sequence of intracellular

IL-1ra type I not in common"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

CAGAAGACCT CCTGTCCTAT GAGGCCCTCC CC ATG GCT TTA G

Met Ala Leu

42

1

- (2) INFORMATION FOR SEQ ID NO: 7:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

Met Ala Leu

1

- (2) INFORMATION FOR SEQ ID NO: 8:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 105 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLE	CULE T	YPE:	CDNA									
(iii) HYF	OTHETI	CAL:	NO									
(iv) ANT	-SENSE	: NO										
(ix) FEAT	URE:											
	(A)	NAME/	KEY:	CDS									
	(B)	LOCAT	ION:	33104	1								
(ix) FEAT	TURE:											
	(A)	NAME/	KEY:	misc_:	featu	ıre							
	(B)	LOCAT	ION:	1105									
	(D)	OTHER	INF	ORMATIO	ON : /r	not e =	* "Se	equer	ice o	of ir	ntrac	cellu	lar
		•				IL-I	ra t	ype	II r	not i	in co	mmon	Ħ
	xi) SEQ											•	
CAGAAGA	CCT CCT	GTCCTAT	GAG	GCCCTC	c cc	ATG	GCT	TTA	GCT	GAC	TTG	TAT	53
						Met	Ala	Leu	Ala	Asp	Leu	Tyr	
						1				5			
	GGA GG												101
Glu Glu	Gly Gl	y Gly G	Sly G	ly Gly	Glu	Gly	Glu	Asp	Asn	Ala	Asp	Ser	
	10			15					20				
AAG G													105
Lys													
	FORMATIO												
•	(i) SEQU												
	-) LENGT				ids							
) TYPE:											
	(D) TOPOI	LOGY :	linea	r								
	(ii) MOL			•				_					
	(xi) SEC								7 1	G1	. Cl.	C1	
Met Ala	a Leu Al		Leu 7	fyr Glu	Glu			GIY	GIY	GIY			
1		5				10					15		
Gly Gl	u Asp As		Asp S	Ser Lys	5								
		0											
(2) IN	FORMATIO												
	(i) SEQU	IENCE C	HARA	TERIS:	TICS:								
		\\ LENG											

(B)	TYPE: nucleic	acıa
(C)	STRANDEDNESS:	single

(6, 51,41,525,500, 51,31

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..468

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1..474
- (D) OTHER INFORMATION:/note= "Common IL-1ra seq.; a G was added in the first position for software reason, so as the first codon codes for Glu and so as the creation of a stop codon in the inner region of the seq. is avoided"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

 GAG ACG ATC TGC CGA CCC TCT GGG AGA AAA TCC AGC AAG ATG CAA GCC 48

 Glu Thr Ile Cys Arg Pro Ser Gly Arg Lys Ser Ser Lys Met Gln Ala

 1 5 10 15
- TTC AGA ATC TGG GAT GTT AAC CAG AAG ACC TTC TAT CTG AGG AAC AAC

 Phe Arg Ile Trp Asp Val Asn Gln Lys Thr Phe Tyr Leu Arg Asn Asn

 20 25 30

CAA CTA GTT GCT GGA TAC TTG CAA GGA CCA AAT GTC AAT TTA GAA GAA 144 Gln Leu Val Ala Gly Tyr Leu Gln Gly Pro Asn Val Asn Leu Glu Glu

35 40 45

AAG ATA GAT GTG GTA CCC ATT GAG CCT CAT GCT CTG TTC TTG GGA ATC 192 Lys Ile Asp Val Val Pro Ile Glu Pro His Ala Leu Phe Leu Gly Ile

50 55 60

CAT GGA GGG AAG ATG TGC CTG TCC TGT GTC AAG TCT GGT GAT GAG ACC

240

His Gly Gly Lys Met Cys Leu Ser Cys Val Lys Ser Gly Asp Glu Thr

65 70 75 80

AGA CTC CAG CTG GAG GCA GTT AAC ATC ACT GAC CTG AGC GAG AAC AGA 288
Arg Leu Gln Leu Glu Ala Val Asn Ile Thr Asp Leu Ser Glu Asn Arg

-20-

				85					90					95		
AAG	CAG	GAC	AAG	CGC	TTC	GCC	TTC	ATC	CGC	TCA	GAC	AGT	GGC	CCC	ACC	336
Lys	Gln	Asp	Lys	Arg	Phe	Ala	Phe	Ile	Arg	Ser	Asp	Ser	Gly	Pro	Thr	
			100					105					110			
ACC	AGT	TTT	GAG	TĊT	GCC	GCC	TGC	CCC	GGT	TGG	TTC	CTC	TGC	ACA	GCG	384
Thr	Ser	Phe	Glu	Ser	Ala	Ala	Cys	Pro	Gly	Trp	Phe	Leu	Cys	Thr	Ala	
		115					120					125				
ATG	GAA	GCT	GAC	CAG	CCC	GTC	AGC	CTC	ACC	TAA	ATG	CCT	GAC	GAA	GGC	432
Met	Glu	Ala	Asp	Gln	Pro	Val	Ser	Leu	Thr	Asn	Met	Pro	Asp	Glu	Gly	
	130					135					140					
GTC	ATG	GTC	ACC	AAA	TTC	TAC	TTC	CAG	GAG	GAC	GAG	TAG	ΓAC		474	
Val	Met	Val	Thr	Lys	Phe	Tyr	Phe	Gln	Glu	Asp	Glu					
145																
(2)	INFO	ORMAT	CION	FOR	SEQ	ID N	10: 1	11:								
	i)	i) SE	EQUE	1CE (CHAR	ACTE	RIST	CS:								
			(A)	LENG	GTH:	156	amin	no ac	cids							
			(B)	TYP	E: ar	nino	acio	3								
					DLOG!											
•					TYPE	_										
	•		_		DES							7	N4	C1-	3 .1.5	
	Thr	He	Cys	_	Pro	Ser	GIY	Arg		Ser	ser	Lys	met		Ald	
1	_		_	5			61 -	.	10	D.b	7 2	T	>	15	200	
Phe	Arg	Ile	_	Asp	Val	Asn	Gin		Inr	Pne	lyr	reu		ASII	ASII	
O1 -	7	17-7	20	G1	Tyr	T 011	C1 n	25	Dro	λαη	Val	λεπ	30	Glu	Glu	
GIII	Leu	35	Ald	GIY	TYL	nea	40	Gry	FIO	ASII	Vai	45	neu	010	Olu	
Lvc	Tlo		Ma l	Val	Pro	Tle		Pro	Hic	Δla	ī.e.ı		Leu	Glv	Tle	
пуъ	50	Asp	Vai	Vai	110	55	010	110	1115	niu	60	1110	Dea	Cly		
uic		Cly	Lvc	Mo+	Cys		Ser	Cvs	Val	Lvs		Glv	Asn	Glu	Thr	
	Gry	Gly	Lys	MEC	70	Dea	361	СуЗ	VUI	75	561	Cly	пор	010	80	
65	Lov	Cln	Len	Glu	Ala	Val	λεη	Tle	Thr		ī.eu	Ser	Glu	Asn		
Ar 9	Leu	3111	₩ eu	85	AIG	VU1	7511	110	90	p				95	- •- 9	
Lvc	Gla	A ~ ~	Tare		Phe	בום	Phe	116		Ser	Asn	Ser	Glv		Thr	
PAR	GIII	vsh	Lys	νιά	: 116	AIG	1110	105		561			110			

-21-

- (2) INFORMATION FOR SEQ ID NO: 12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS:
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (v) FRAGMENT TYPE: N-terminal
 - (ix) FEATURE:
 - (A) NAME/KEY: Peptide
 - (B) LOCATION:1..21
 - (D) OTHER INFORMATION:/note= "A portion of the

10

15

intracellular IL-1ra type II not in common"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

Ala Asp Leu Tyr Glu Glu Gly Gly Gly Gly Gly Gly Gly Glu Asp

Asn Ala Asp Ser Lys

20

- (2) INFORMATION FOR SEQ ID NO: 13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 579 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA
 - (iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

	(i	x) F	EATU	RE:												
			(A)	NAME	/KEY	: CD	S									
			(B)	LOCA	TION	:34.	.573									
	(i	x) F	EATU	RE:												
			(A)	NAME	/KEY	: mi	sc_f	eatu	re							
			(B)	LOCA	MOIT	1:1	579									
			(D)	ОТНЕ	RIN	FORM	ATIC	N:/r	ote=	"In	trac	ellu	lar	IL-1	ra t	ype
												II"				
	(x	i) S	EQUE	NCE	DESC	RIPT	: NOI	SEC	OI C	NO:	13:					
CAGA	AGGA	CC T	CCTG	TCCT	A TO	AGGC	CCTC	ccc	ATG	GCT	TTA	GCT	GAC	TTG	TAT	54
									Met	Ala	Leu	Ala	Asp	Leu	Tyr	
									. 1	•			5	5		
GAA	GAA	GGA	GGT	GGA	GGA	GGA	GGA	GAA	GGT	GAA	GAC	AAT	GCT	GAC	TCA	102
Glu	Glu	Gly	Gly	Gly	Gly	Gly	Gly	Glu	Gly	Glu	Asp	Asn	Ala	Asp	Ser	
		10					15					20				
						CCC										150
Lys	Glu	Thr	Ile	Cys	Arg	Pro	Ser	Gly	Arg	Lys	Ser	Ser	Lys	Met	Gln	
	25					30					35					
						GTT										198
Ala	Phe	Arg	Ile	Trp	Asp	Val	Asn	Gln	Lys	Thr	Phe	Tyr	Leu	Arg	Asn	
40					45					50					55	
						TAC										246
Asn	Gln	Leu	Val	Ala	Gly	Tyr	Leu	Gln	Gly	Pro	Asn	Val	Asn		Glu	
				60					65					70		
						CCC										294
Glu	Lys	Ile	Asp	Val	Val	Pro	Ile	Glu	Pro	His	Ala	Leu		Leu	GIY	
			75					80					85			2.4.5
						TGC										342
Ile	His	Gly	Gly	Lys	Met	Cys			Cys	Val	Lys		GIY	Asp	GIU	
		90					95					100		a		3.00
						GCA										390
Thr	Arg	Leu	Gln	Leu	Glu	Ala	Val	Asn	Ile	Thr			Ser	Glu	ASN	
	105					110					115					

-23**-**

	220	CNC	CNC	אממ	cac	TTC	GCC	TTC	ΔΤΟ	CGC	TCA	GAC	AGT	GGC	CCC		438
														Gly			
	Lys	GIII	ASP	гуз		FILE	AIG	FIIC	110	130	561	p	001	017	135		
120		».Com	4	CNC	125 TCT	ccc	ccc	TCC	CCC		тсс	TTC	СТС	TGC			486
Thr	Thr	Ser	Pne		Ser	Ala	Ala	Cys		GIY	пр	FILE	Leu	Cys	1111		
				140					145	1.00	3 A TT	N.T.C	CCT	150	C		534
														GAC			734
Ala	Met	Glu		Asp	GIn	Pro	vai		rea	IIII	ASII	Mec		Asp	GIU		
			155				m> 0	160	CAC	CNC	CAC	CAC	165	EA.C			579
			GTC										IAG.	IAC			3/3
Gly	Val		Val	Thr	rys	Pne		Pne	GIN	GIU	Asp						
		170					175					180					
(2)			иоіл														
	(:	L) SE	EQUE														
								no ac	cids								
			•				acio										
							inear										
			OLE														
			SEQUI									~ .	~ \	61	a.		
Met	Ala	Leu	Ala		Leu	Tyr	Glu	Glu		Gly	GIY	GIÀ	GIY	Gly	GIU		
1				5					10		_		. .	15	G 3		
Gly	Glu	Asp	Asn	Ala	Asp	Ser	Lys		Thr	Ile	Cys	Arg		Ser	GIY		
			20					25					30				
Arg	Lys	Ser	Ser	Lys	Met	Gln	Ala	Phe	Arg	Ile	Trp		Val	Asn	Gln	:	
		35					40					45					
Lys	Thr	Phe	Tyr	Leu	Arg	Asn	Asn	Gln	Leu	Val	Ala	Gly	Tyr	Leu	Gln		
	50					55					60						
Gly	Pro	Asn	Val	Asn	Leu	Glu	Glu	Lys	Ile	Asp	Val	Val	Pro	Ile	Glu		
65					70					75					80		
Pro	His	Ala	Leu	Phe	Leu	Gly	Ile	His	Gly	Gly	Lys	Met	Cys	Leu	Ser		
				85					90					95			
Cys	Val	Lys	Ser	Gly	Asp	Glu	Thr	Arg	Leu	Gln	Leu	Glu	Ala	Val	Asn		
			100					105					110				
Tle	Thr) CD	Len	Sar	Glu	Asn	Ara	Lvs	Gln	Asp	Lvs	Ara	Phe	Ala	Phe		

-24-

		115					120					125			
Ile	Arg	Şer	Asp	Ser	Gly	Pro	Thr	Thr	Ser	Phe	Glu	Ser	Ala	Ala	Cys
	130					135					140				
Pro	Gly	Trp	Phe	Leu	Cys	Thr	Ala	Met	Glu	Ala	Asp	Gln	Pro	Val	Ser
145					150					155					160
Leu	Thr	Asn	Met	Pro	Asp	Glu	Gly	Val	Met	Val	Thr	Lys	Phe	Tyr	Phe
				165					170					175	
Gln	Glu	Asp	Glu												
			180												

10

15

30

-25-

CLAIMS

- 1. Purified protein having antagonist activity against at least one of the substances selected from the group consisting of interleukin a and interleukin B, or fragments thereof, characterised by the fact of comprising the amino acid sequence reported in SEQ ID NO:12.
- 2. A purified protein having antagonist activity against at least one of the substances selected from the group consisting of interleukin a and interleukin B, or fragments thereof, characterised by the fact to have the amino acid sequence reported in SEQ ID NO:14.
- 3. A purified protein according to claim 2, characterised by the fact of being obtained by recombinant DNA technique.
- 4. Isolated DNA sequence encoding a IL-1 antagonist having the amino acid sequence according to claim 2.
- 5. Isolated DNA sequence according to claim 4, characterised by the fact of having reported in SEQ ID NO:13.
 - 6. Isolated DNA sequence that ibridizes with the probe having the sequence reported in SEQ ID NO:1.
- 7. Vector comprising the DNA sequence encoding the protein according to claim 2.
 - 8. Cell culture transfected with DNA encoding the protein according to claim 2.
 - 9. Cell culture according to claim 2, characterised by the fact of being obtained by mammalian cells.
- 10. Process for obtaining a IL-1 antagonist by the recombinant DNA technique comprising:
 - a) cultivating host cells as described in claims 8 and 9 containing a DNA sequence able to produce the protein according to claim 2;

- b) collecting and isolating the encoded protein.
- 11. Process of obtaining a IL-1 antagonist by recombinant DNA technique. characterised by the fact that the DNA sequence is that reported in SEQ ID NO:13.
- 12. Use of a purified protein having the protein according to claim 2, for medical use.
- 13. Use of a purified protein having the sequence according to claim 2 for the preparation of pharmaceutical compositions active in pathologies requiring IL-1 inhibition.
- 14. Use according to claim 13, characterised by the fact that the pathology is selected from the group of autoimmune pathologies.
- 15. Use according to claim 14, characterised by the fact that the pathology is selected from the group consisting of rheumatoid arthritis, septic shock, acute myelomonocytic leukaemia, immunological reaction of transplantation against host, acquired immunodeficiency syndrome (AIDS), ulcerative colitis.

Figure 1

Secreted IL-1ra

Ö TCA Met Glu Ile Cys Arg Gly Leu Arg Ser His Leu Ile Thr Leu Leu Leu Phe Leu Phe His Ser CAT TTC TTC CTG CTA ATC ACT CTC CTC CTC CTC CGC AGT CAC GAATTCCGGG CTGCAGTCAC AGA ATG GAA ATC TGC AGA GGC

Intracellular IL-1ra type I

AXI AXI CAGAAGACCT CCTGTCCTAT GAGGCCCTCC CC ATG GCT TTA

Met Ala Leu

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Intracellular IL-1ra type II

IRA

IRA 5

Glu Met Ala Leu Ala Asp Leu Tyr Glu Glu Gly Gly Gly Gly Gly Gly

1/5

GAA GAC AAT GCT GAC TCA AAG G Glu Asp Asn Ala Asp Ser Lys

Common IL-1ra sequence

Thr Lys CAG AAG Asn Gln AAC GTT Trp Asp Val TGG GAT ATC Ile Arg AGA TTCPhe CCC Ala Gln ATG CAA Met Lys AGC AAG Ser Ser AGA AAA TCC Lys Arg 999 Gly AG ACG ATC TGC CGA CCC TCT Pro Ser Arg Cys I1e

Val GAT GAA AAG ATA Ile Lys Glu Glu TTA GAA Leu AAT Asn GTC Val Asn CCA AAT Pro CAA GGA Gln Gly TTG Leu TAC Tyr GGA Gly GCT Ala Val CTA GTT Leu CAA Gln AAC AAC Asn Asn AGG Arg CTG Leu TAT TTC

GAT GGTG1ySer GTC AAG TCT Cys Val Lys TGT TCCSer CTGLeú Cys TGC ATG Ala Leu Phe Leu Gly Ile His Gly Gly Lys Met AAG GGA GGG CAT GGA ATC TTGTTCCTGGCT CAT Glu Pro His CCTGAG ATT Val Pro Ile

Figure 1 cont.

Glu Ala Asp Gln Pro Val Ser Leu Thr Asn Met Pro Asp Glu Gly Val Met Val Thr Lys Phe Tyr Phe Gln Glu Asp Glu TTC GAA GCT GAC CAG CCC GTC AGC CTC ACC AAT ATG CCT GAC GAA GGC GTC ATG GTC ACC AAA TTC TAC TTC CAG GAG GAC GAG Glu Thr Arg Leu Gln Leu Glu Ala Val Asn Ile Thr Asp Leu Ser Glu Asn Arg Lys Gln Asp Lys Arg Phe Ala GAG TCT GCC GCC TGC CCC GGT TGG TTC CTC TGC ACA GCG ATG GAG ACC AGA CTC CAG CTG GAG GCA GTT AAC ATC ACT GAC CTG AGC GAG AAC AGA AAG CAG GAC AAG CGC TTC GCC ATC CGC TCA GAC AGT GGC CCC ACC ACC AGT TTT GAG TCT GCC GCC TGC CCC GGT TGG TTC CTC TGC ACA GCG ATG Ile Arg Ser Asp Ser Gly Pro Thr Thr Ser Phe Glu Ser Ala Ala Cys Pro Gly Trp Phe Leu Cys Thr Ala Met

TAGTAC

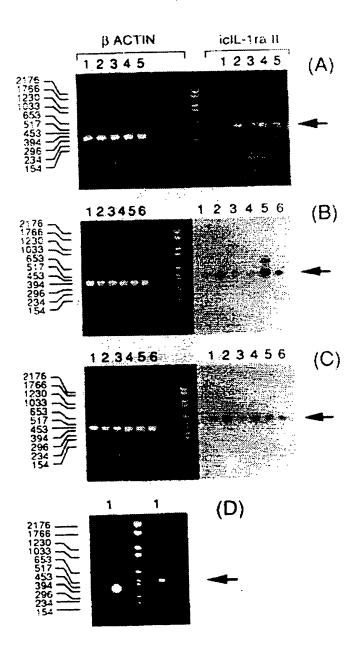
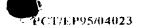


Figure 2



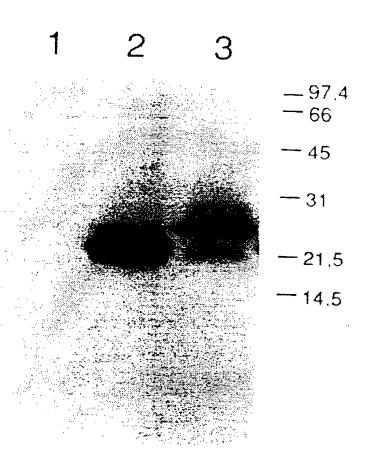


Figure 3

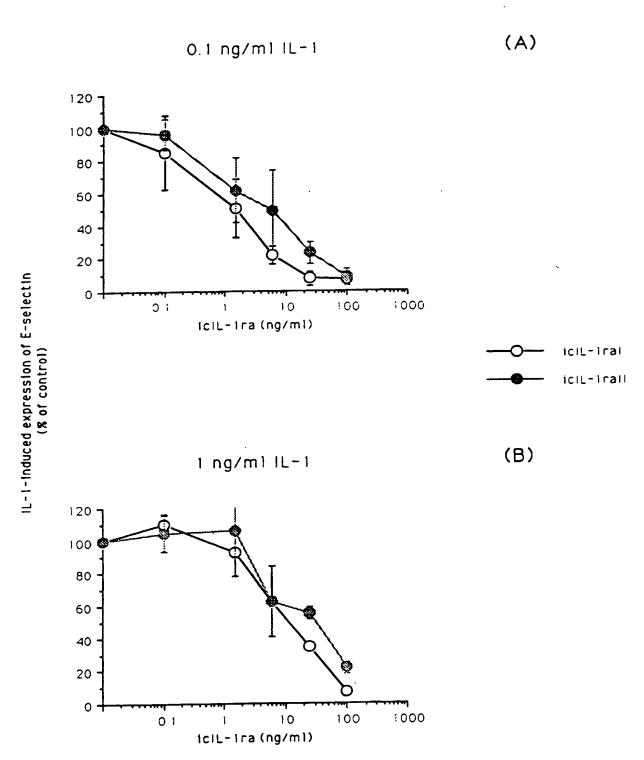


Figure 4

INTERNATIONAL SEARCH REPORT

ication No PCT/EP=95/04023

A. CLASSIFICATION OF SUBJECT MATTER 1PC 6 C12N15/24 C07K14/54 A61K38/20 C12N5/10 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) C07K C12N IPC 6 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Relevant to claim No. Citation of document, with indication, where appropriate, of the relevant passages Category ' 1-11 PROCEEDINGS OF THE NATIONAL ACADEMY OF Α SCIENCES OF USA, vol. 88, 1991 WASHINGTON US, pages 3681-3685, S. HASKILL ET AL 'cDNA cloning of an intracellular form of the human Interleukin 1 receptor antagonist associated with epithelium' cited in the application see the whole document -/--Patent family members are listed in annex. Further documents are listed in the continuation of box C. X Special categories of cited documents: "T" later document published after the international filing date or priority date and not in conflict with the application but "A" document defining the general state of the art which is not ated to understand the principle or theory underlying the considered to be of particular relevance invention 'E' earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-'O' document referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person stilled other means document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of mailing of the international search report Date of the actual completion of the international search 15.03.96 28 February 1996 Authorized officer Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Ripwik Tel. (+31-70) 340-2040, Tx. 31 651 epo ni,

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Fax (+31-70) 340-3016

Le Cornec, N



rational Application No PCT/EP 95/04023

		PC1/EP 95/04023
	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	10
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	JOURNAL OF IMMUNOLOGY, vol. 153, July 1994 BALTIMORE US, pages 701-711, C. BUTCHER ET AL 'Comparison of two promoters controlling expression of secreted or intracellular il-1 receptor antagonist' see the whole document	1-12
A	IMMUNOLOGY TODAY, vol. 12, no. 11, 1991 CAMBRIDGE GB, pages 404-410, C. A. DINARELLO ET AL 'Blocking il-1: Interleukin 1 receptor antagonist in vivo and in vitro' see the whole document	1,2, 12-15
X	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 85, May 1988 WASHINGTON US, pages 2929-2933, N. F. ZANDER ET AL 'cDNA cloning and complete primary structure of skeletal muscle phosphorylase kinase(alpha subunit)' see figure 2	6
X	EMBL database entry SKADECYC Accession number X56042 (version 1);19 november 1990; YOUNG D. et al see abstract	6
A .	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 88, June 1991 WASHINGTON US, pages 5232-5236, S.P. EISENBERG ET AL 'INterleukin 1 receptor antagonist is a member of the interleukin 1 gene family: Evolution of a cytokine control mechanism'	
A	WO,A,91 17249 (CETUS CORPORATION) 14 November 1991 see abstract; claims; table I see page 29; figure 2	1-15
P,X	JOURNAL OF EXPERIMENTAL MEDICINE, vol. 182, no. 2, 1 August 1995 pages 623-628, M. MUZIO ET AL 'Cloning and characterization of a new isoform of the interleukin 1 receptor antagonist' see the whole document	1-12

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Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
	rnational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
i. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Remark: Although claim 12 is directed to a method of treatment of the human/animal body (rule 39.1(iv)PCT) the search has been carried out and based on the alleged effects of the compound/composition.
2.	Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box il	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Int	ternational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.	As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4.	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remar	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

nformation on patent family members

:mational Application No PCT/EP 95/04023

Publication date	Patent memb		Publicauon dale
14-11-91	AU-B-	655766	12-01-95 27-11-91
	CA-A-	2081774	02-11-91
	EP-A-	0534978	07-04-93
	US-A-	5455330	03-10-95
	date	14-11-91 AU-B- AU-B- CA-A-	14-11-91 AU-B- 655766 AU-B- 7692791 CA-A- 2081774 EP-A- 0534978